**Supplementary File 1**

**Chemicals/reagents and equipment**

The chemicals/reagents used for the study were of the best grade available. All the chemicals/reagents and the equipment used are listed in the **Table of Materials**.

1. **Animal experiment**

8 weeks old C57BL6 mice were used in this study. Briefly, *P. gingivalis* (ATCC 33277) was cultured in anaerobic conditions in Brain heart Infusion medium supplemented with hemine and menadione as described previously. 5 x 107 bacteria were injected intra-peritoneally (IP) 3 times a week to each animal for 2 weeks. All experimental protocols were approved by the Ethics Committee of Strasbourg named “Comité Régional d’Ethique en Matière d’Expérimentation Animale de Strasbourg (CREMEAS)” (APAFIS#28745-2020121815051557) and followed relevant guidelines of the same. Mice were examined to evaluate pain and stress, and their weights were monitored daily.

Animal’s sacrifice was performed after anesthesia using a combination of 100 mg/kg of ketamine and 10 mg/kg of xylazine, and then the spleen was harvested after laparotomy.

1. **Quantification of splenocyte microvesicles by prothrombinase enzymatic assay**

This assay will measure the MVs that expose anionic phospholipids (mainly PS). In this enzymatic assay, the degree of exposure of PS on the surface of MVs is the limiting factor of the assembly reaction of the prothrombinase coagulation complex, which leads to the release of soluble thrombin from prothrombin in the presence of factor Xa and cofactor Va. Thrombin is revealed using the chromogenic substrate pNAPEP. The measurements are carried out using a multiwell plate thermostated photometer equipped with kinetics software in a standardized reaction medium containing 2.3 mM free calcium and highly purified human coagulation factors. Absorbances are converted into PS equivalents (nM PS eq.) by reference to a calibration curve obtained using a suspension of synthetic vesicles containing known amounts of PS and PtdChol 33. Such a calibration offers the advantage of measuring phosphatidylserine as a catalytic surface, regardless of an eventual difference in the amount of phosphatidylserine exposed from one MV to another. Therefore, should the proportion of phosphatidylserine vary from one MP to another, the normalization to a total phosphatidylserine membrane surface allows comparison between samples. MP concentrations are referred to as nanomolar phosphatidylserine “equivalents”.

In this assay, mean repeatability measured with standard synthetic vesicle concentrations or washed MVs concentrations is 10.3 %. A 2.1 nM concentration from the lowest standard vesicle curve can be measured with a 10% precision, and 0.5 nM is the lowest detectable value.

1. **Spectrophotometric measurement of splenic microvesicles proteins and RNA concentration by NanoDrop**
   1. Select the “Proteins tab” and press **Protein A280** and/or **A260** for RNA measurement from the home screen.
   2. Specify a type of sample.
   3. Pipet 1–2 μL of control solution (HBSS), and place on the lower pedestal and lower the arm
   4. Press **Blank** and wait for the measurement to be completed.
   5. Lift the arm and clean the two bases with a new laboratory wipe of optical grade or remove the stopper cuvette.
   6. Pipette 2 μL of sample solution (don’t forget to mix thoroughly) place on the pedestal and lower the arm, insert the sample cell into the cell holder.
   7. Start the measurement.
   8. Press **End** of the experiment once the measuring of the samples is over.
   9. Lift the arm and clean the two bases with a new wipe or remove the sample cell.

NOTE: Get 3 typical data for the quantification of MVs: The first is a rough evaluation of the protein content of the MVs (mg/mL), the second for nucleic acids, and the third for the corresponding purity ratios. In our hands, the correlation curve can be drawn between MV concentration measured as a protein concentration using protein colorimetric assay of MV lysates and a calibrated albumin reference curve, and spectrophotometry (nanodrop) using intact MVs. Once the correlation is established under each specific experimental condition (control MVs, MVs from stimulated cells or animals), the nanodrop measurement can be a valuable alternate tool to save sample volume and time. Another quantification procedure is the prothrombinase assay (see below).

1. **Determination of the proportion of neutrophils in the splenocyte suspension** 
   1. In a 2 mL microcentrifuge tube, put 1 x 106 cells.
   2. Centrifuge the tube for 1 min at 132 x *g* at room temperature and throw the supernatant.
   3. Add 1x PBS to the pellet and wash twice using two centrifugation steps.
   4. Prepare antibodies cocktail dilutions in 5% BSA solution:

CD11b specific of Monocytes, neutrophils, granulocytes: (1/50)

YG-6 Gr1 specific for granulocytes: (1/200)

LY6-C specific for neutrophils: (1/200)

* 1. Block the splenocytes with 50 μL of FC block (CD16/CD32) for 1 h on ice to avoid unspecific binding of Fc fragments of IgGs to their CD16 /CD32 receptor (1μg/sample).
  2. Wash twice with prepared 1x PBS.
  3. Add 50 μL of the antibodies cocktail into the microcentrifuge tube and incubate in the dark for 15 min on ice.
  4. Wash twice with prepared 1x PBS.
  5. Fill and resuspend the pellet with 1x PBS (300 μL).
  6. Transfer into specific tubes for cytometry across a 0.1 μm filter.
  7. Define the gate of living using DAPI fluorescence. Construct a scatter plot with LIG-6 Gr-1 and CD11b (Mac-1). Neutrophils are delineated as Gr1+ / Mac-1+. Flow cytometric compensation was performed using single labeling.
  8. Cells were analyzed using multiparameter flow cytometry. The fluorescence minus one (FMO) principle was utilized to identify positive and negative populations to account for background antibody fluorescence.
  9. Pass each sample into the flow cytometer.
  10. Analyze using Flow Jo software or the software proposed by the cytometer manufacturer.

NOTE: Control-stained cells can be stored at 4 °C under sterile conditions in case of a second acquisition at t0 + 24 h. Data shown in **Figure 5** were obtained using freshly isolated stained cells. However, the cell fixation procedure is to be established for antibodies affinity and cell viability.

Neutrophil counts are determined into the gate delineated for LY6-G positive and CD11b positive, LY6C low stained cells. In addition, preliminary cell count using DAPI will discriminate all dead cells. Typically, 10,000 cells are sufficient for acquisition and accurate analysis.

**Details on supplementary figures**

**Supplemental Figure 1:** **How procoagulant MVs are procoagulant**

In vascular cells, binding of various agonists to their specific receptor induces calcium influx that will contribute to the swift translocation of phosphatidylserine (PSer), an anionic phospholipid, from the inner leaflet to the outer leaflet under the effect of a marked floppase net activity in response to the influx while the reverse transporters of anionic phospholipids (flippase) become inhibited. PS externalization is more rapid than the reverse outer to inner transport of other non-anionic phospholipids such as phosphatidylcholine. This results in an overload of the outer leaflet and budding of the membrane due to surface tensions and is favored by the activation of cysteine-proteases such as calpains and ROCK kinases by high calcium concentrations leading to controlled proteolysis of the cytoskeleton. MVs are procoagulant because they carry PSer, which catalyzes coagulation reactions and constitutes an additional surface for the assembly of blood coagulation complexes. Scott syndrome patients have a bleeding disorder and are characterized by deficient PSer externalization and MV shedding. MVs are also cellular effectors, eventually contributing to disseminating a procoagulant phenotype by activating vascular cells.

**Supplemental Figure 2: The MP captured-based prothrombinase assay**

Left panel: Biotinylated annexin-5 (yellow rectangles) is insolubilized onto streptavidin-coated microplate wells. MPs are captured by phosphatidylserine binding to annexin-5 (phospholipid yellow polar heads) (Total MPs). Middle panel: After 3 washings in Tris Buffer Saline containing 0.05% Tween 20, insolubilized MPs are rinsed once in the absence of Tween 20, and human purified clotting factors are added at concentrations ensuring that phosphatidylserine is the rate-limiting factor of the prothrombinase activity. Factor Xa converts prothrombin (Factor II) into thrombin in the presence of cofactor FVa and calcium. Right panel: Kinetics of thrombin generation by the prothrombinase complex is assessed through the cleavage of a highly specific chromogenic substrate (PNAPEP) and absorbance recorded at 405 nm.

**Supplemental Figure 5**: **Identification of the protein content of MVs from rat splenocyte MVs isolated from young rats by western blot**

Analysis of ICAm-1 and VCAM-1 enrichment of the SMVs shed by rat splenocytes in response to PMA/ionophore A23187. The MVs were harvested, washed, and concentrated, and then the proteins were extracted by incubation in the RIPA lysis buffer. Identical amounts of protein extracts were submitted to electrophoresis based on a protein concentration colorimetric assay. The histograms represent the cumulative values ​​of 3 experiments using splenocytes freshly isolated from 3 different rats. The optical density values ​​are calculated as a ratio to the density of the housekeeping protein β-actin. CTL MVs of unstimulated splenocyte cells: PMA/I: MVs of splenocytes stimulated with PMA (25 ng/mL)/Ionophore A23187 (1 μM, n = 3, \**p* < 0.05 *vs*. CTL).